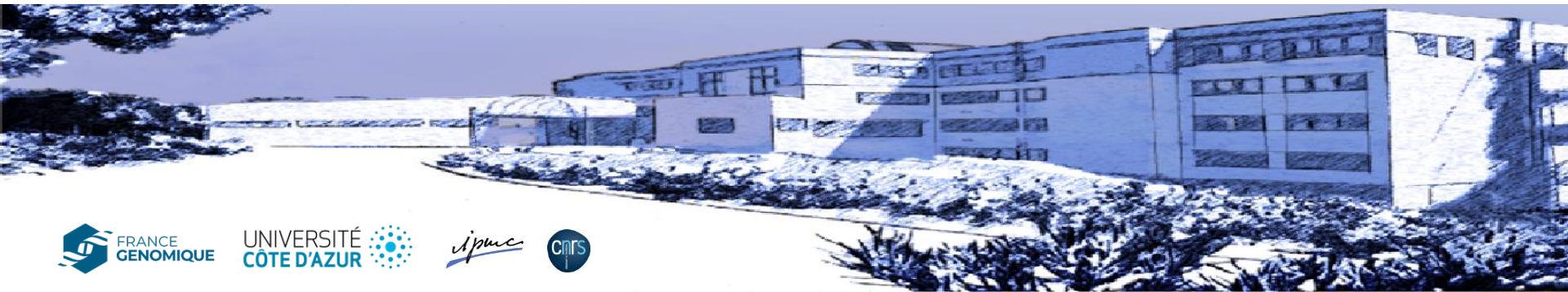


# Single-cell to Spatial Isoform Transcriptomics

Kévin LEBRIGAND, PhD  
Institut de Pharmacologie Moléculaire et Cellulaire  
Université Côte d'Azur, CNRS, France.

✉ [lebrigand@ipmc.cnrs.fr](mailto:lebrigand@ipmc.cnrs.fr)  
🐦 [@kevinlebrigand](https://twitter.com/kevinlebrigand)  
🌐 [@kevinlebrigand@genomic.social](https://www.linkedin.com/company/kevinlebrigand@genomic.social)



# 01

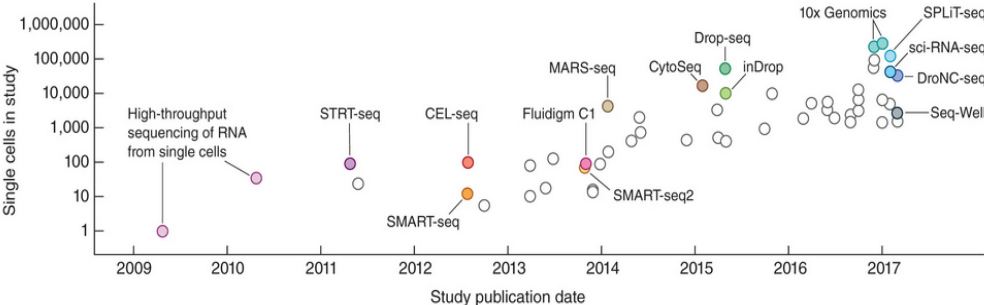
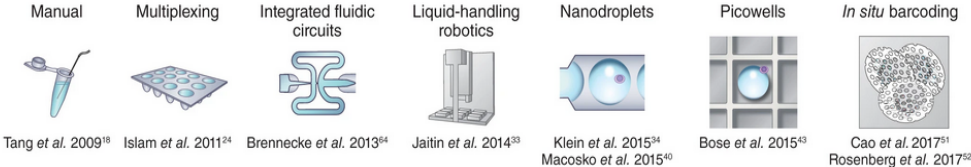
## Single-cell Transcriptomics (NGS)

*Nature Method Of the Year 2013*



# Single-cell transcriptomics

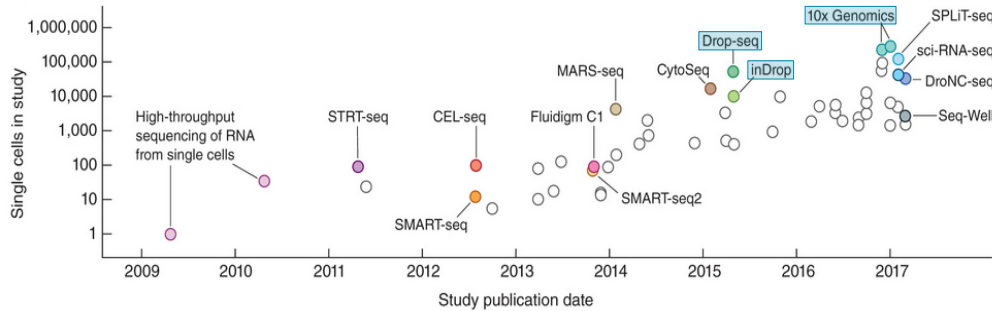
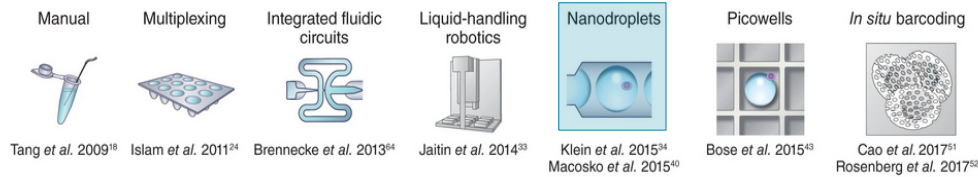
Evolution of isolation techniques and throughput



Exponential scaling of single-cell RNA-seq in the past decade  
 Svensson et al., *Nature Protocols*, 2018

# Single-cell transcriptomics

## Droplet-based approaches

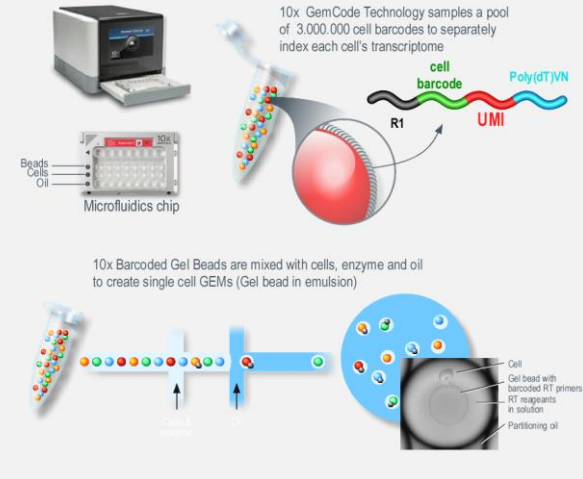


Exponential scaling of single-cell RNA-seq in the past decade  
Svensson et al., *Nature Protocols*, 2018

InDrop, Klein et al, 2015  
Drop-seq, Macosko et al, 2015  
10x Genomics, Zheng et al, 2016

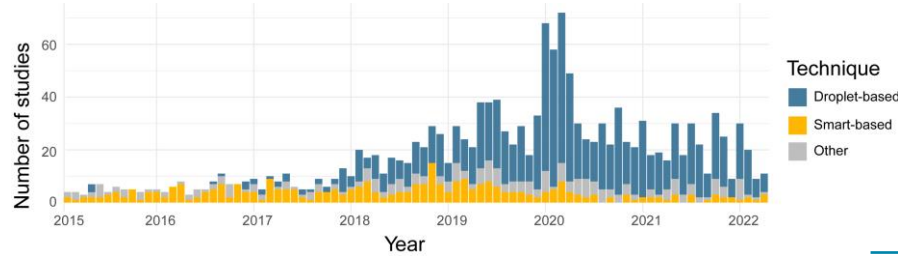
## 10x Genomics Chromium single cell controller (2016)

- Easy-to-set-up and robust workflow
- High scalability (1,3M cells dataset)



# Single-cell transcriptomics

Single cell approaches in publications



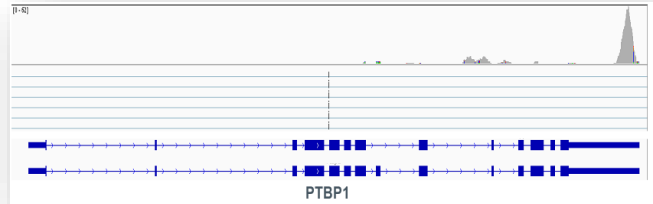
A curated database reveals trends in single cell transcriptomics  
Svensson et al., Database, 2020

- Huge amount of single-cell studies in the past 5 years,
- Droplet-based approaches = 61% (Chromium: 47%)

Technique	Count
Chromium	725
Smart-seq2	177
SMARTer (C1)	124
Drop-seq	74
SMARTer	28
InDrops	23
CITE-seq	18
CEL-seq2	17
STRT-seq	17
MARS-seq	16
Tang	15
CEL-seq	13
STRT-seq (C1)	13
Seq-Well	13
SORT-seq	12
BD Rhapsody	11
BioMark	8
GemCode	7
ICELL8	7
Perturb-seq	7
Patch-seq	6
sc-RT-mPCR	6
MERFISH	5

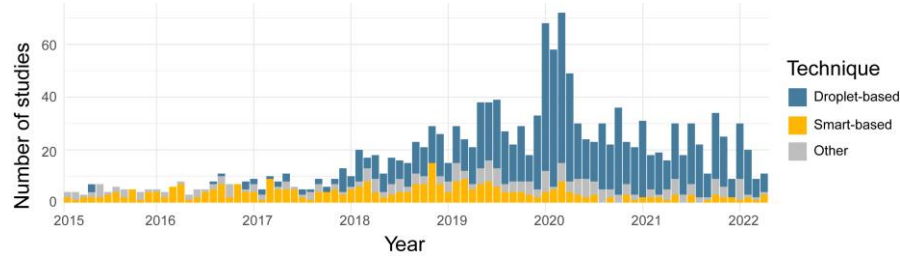
## Droplet-based approaches

- Digital Gene Expression (UMI)
- High cell number throughput
- Limited capture efficiency (<10%)
- 3p or 5p signal (SAGE-like)



# Single-cell transcriptomics

Single cell approaches in publications



A curated database reveals trends in single cell transcriptomics  
Svensson et al., Database , 2020

- Huge amount of single-cell studies in the past 5 years,
- Droplet-based approaches = 61% (Chromium: 47%)
- Smart-based approach = 21%, <5% in the last 2 years

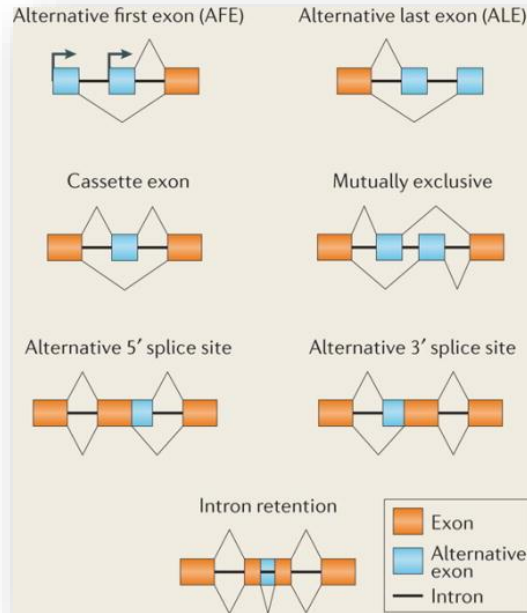
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MARS-seq	16
Tang	15
CEL-seq	13
STRT-seq (C1)	13
Seq-Well	13
SORT-seq	12
BD Rhapsody	11
BioMark	8
GemCode	7
ICELL8	7
Perturb-seq	7
Patch-seq	6
sc-RT-mPCR	6
MERFISH	5

**Smart-based approach**

- No UMI before v3 (may 2020)
- Lower cell number (384-plate handling)
- Higher capture efficiency (~30%)
- Full-length coverage **using short-reads**

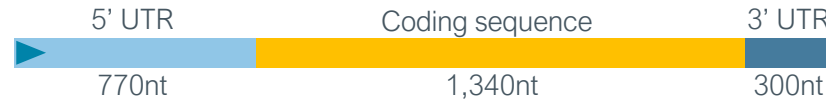
# Transcriptomics

## Complex outcomes of alternative splicing



Scotti and Swanson, Nat Rev Genet., 2016

- 90% of the genes are subjected to alternative splicing,
- Gencode v42 : 252,416 distinct isoforms for 62,696 genes,
- On average, a human gene contains **8.8 exons**, mean size of 145 nt,
- Average encodes mRNA **2,410 nt long** :



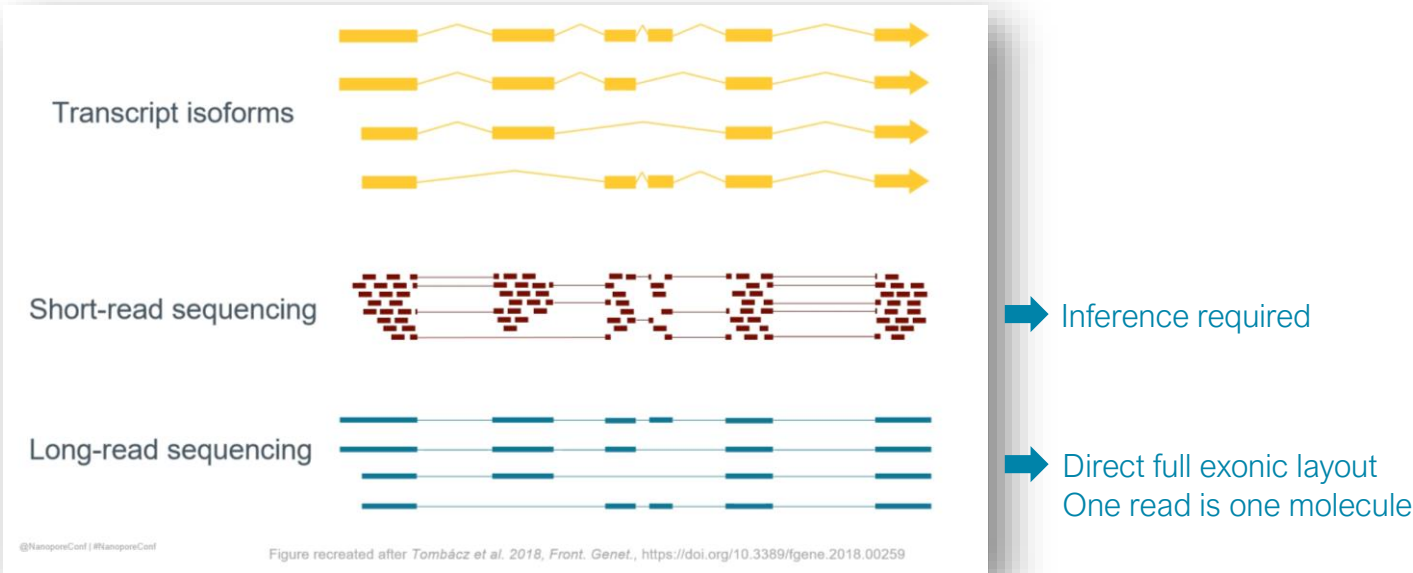
Alternative splicing and disease  
Tazi et al., 2008

# Long-read sequencing identifies isoforms efficiently

Nature Method Of The Year 2022



Nature Method of the Year 2022



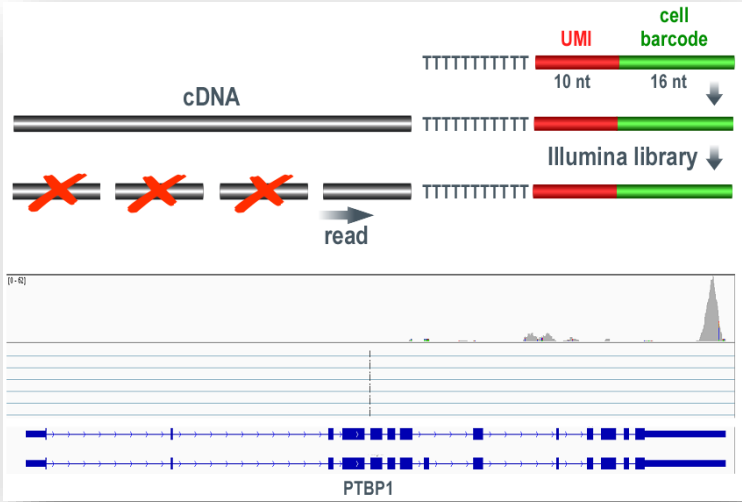


# Single-cell long-read transcriptomics

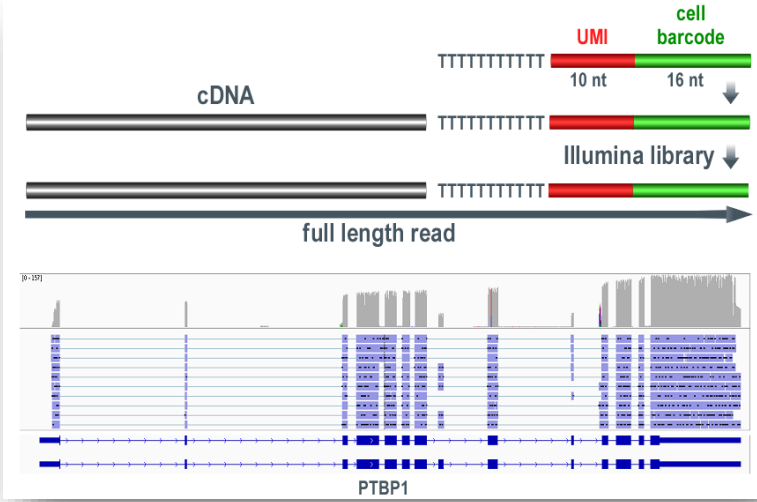
Droplets-based approach short reads vs long reads



Illumina Nextseq500



Nanopore PromethION



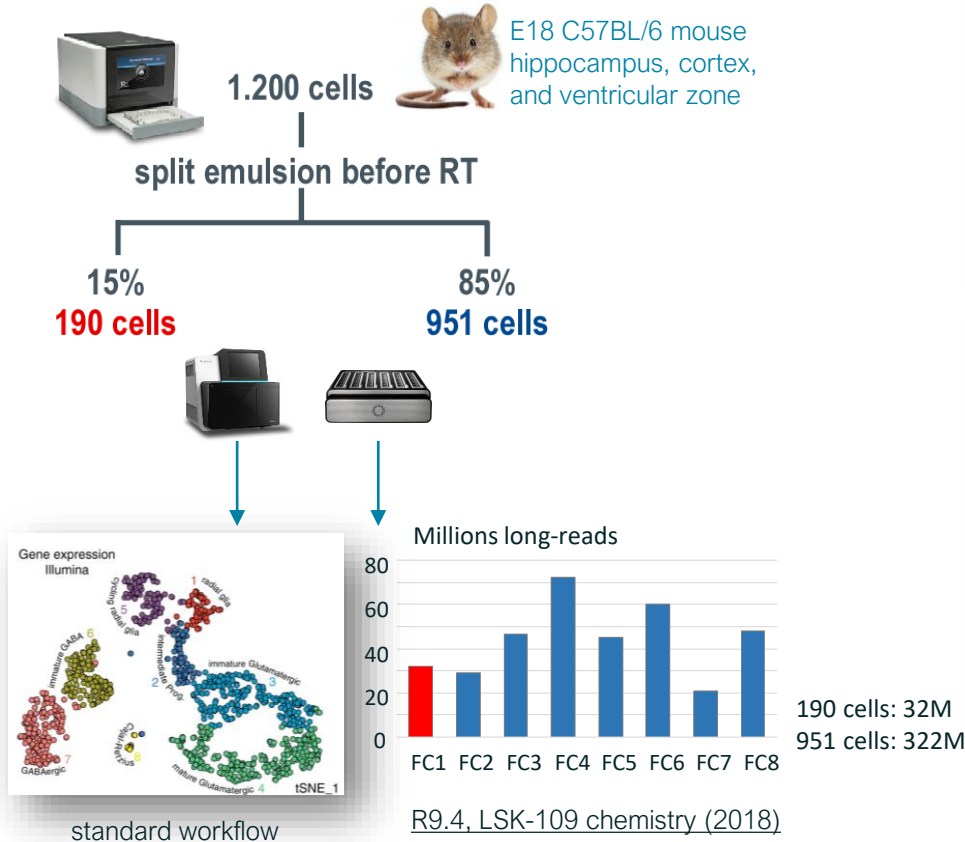
Information on alternative splicing, fusion transcripts, SNV, editing, imprinting

is lost

remain accessible

# Single-cell long-read transcriptomics

SiCeLoRe, bioinformatics for Single Cell Long Read



## nature communications

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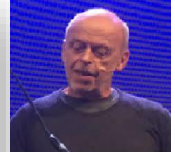
Article | Open Access | Published: 12 August 2020

### High throughput error corrected Nanopore single cell transcriptome sequencing

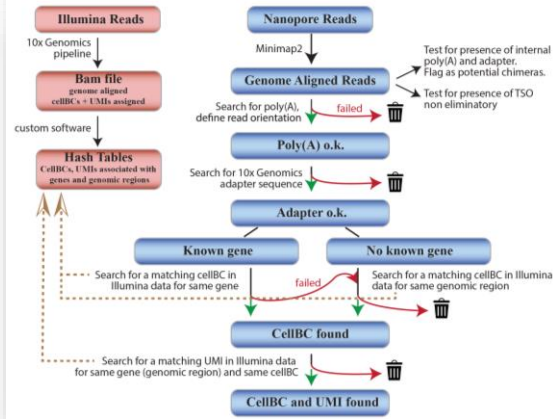
Kevin Lebrigand, Virginie Magnone, Pascal Barby & Rainer Waldmann

Nature Communications 11, Article number: 4025 (2020) | Cite this article

20k Accesses | 38 Citations | 58 Altmetric | Metrics



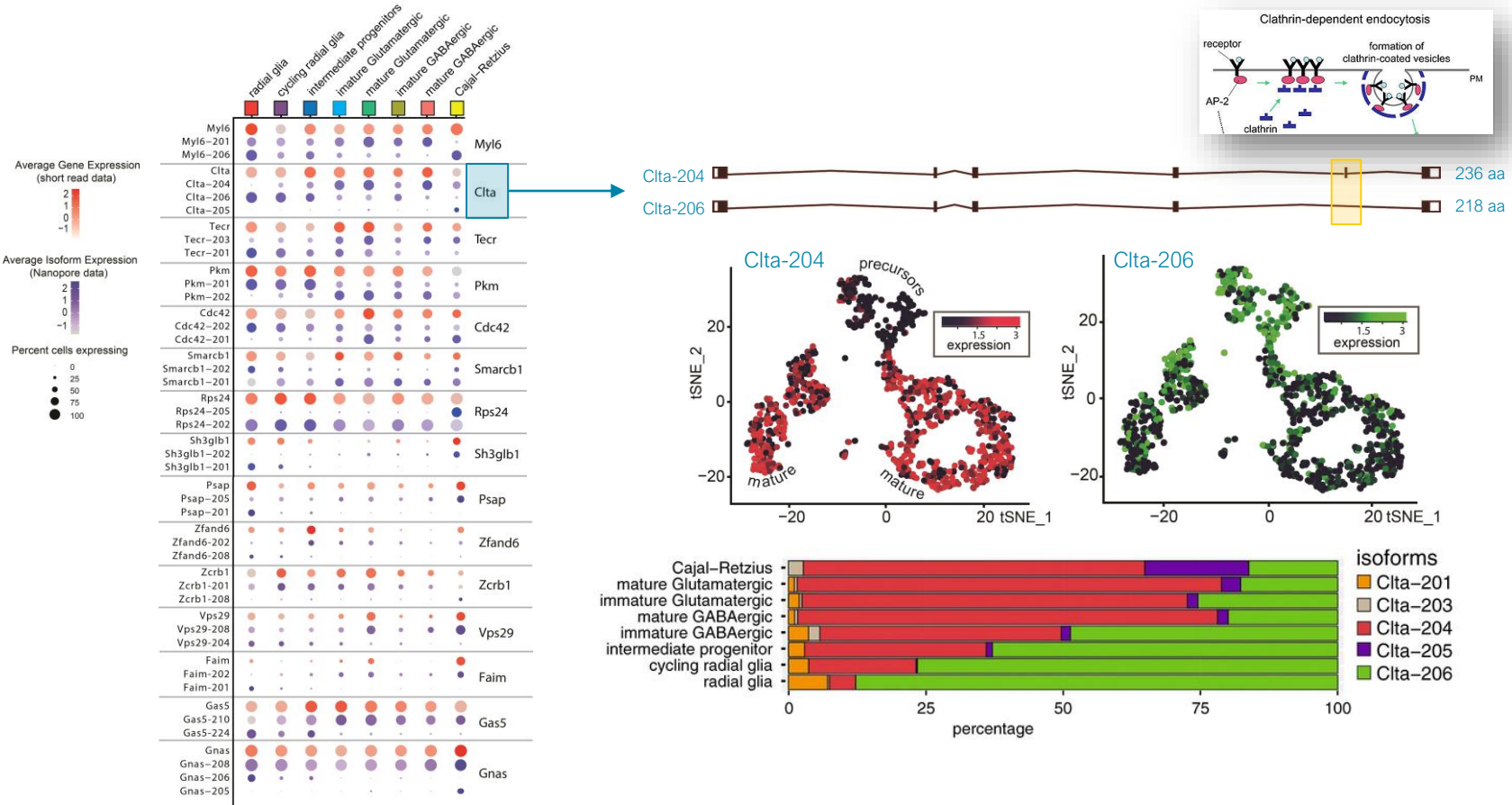
Rainer Waldmann



<https://github.com/ucagenomix/sicelore>

# Single-cell long-read transcriptomics reveals diversity

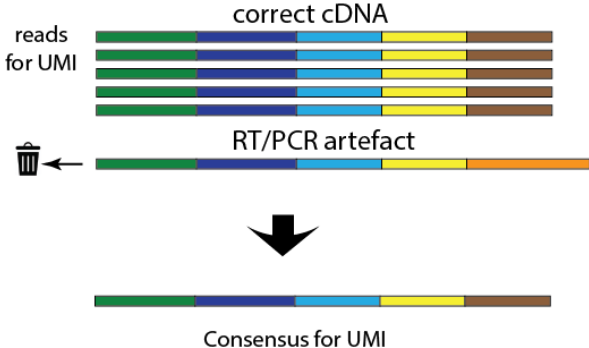
76 isoform-switching genes along neuronal maturation



# Single-cell long-read transcriptomics reveals sequence heterogeneity

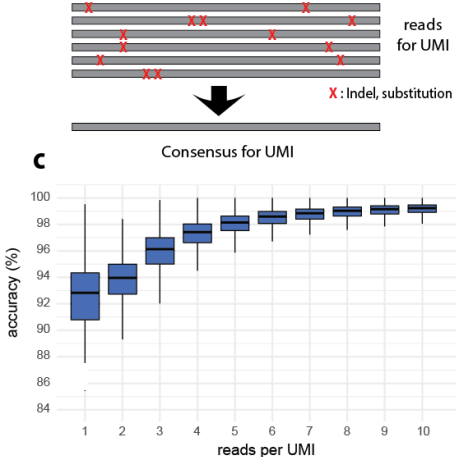
Consensus sequence computation per UMI

UMIs enable elimination of PCR artifacts



Crucial for accurate novel isoform discovery

UMIs enable correction of sequencing errors

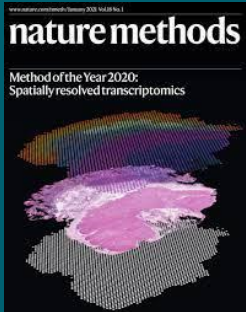


High accuracy for Single Nucleotide Variation call

# 02

## Spatial Transcriptomics (NGS)

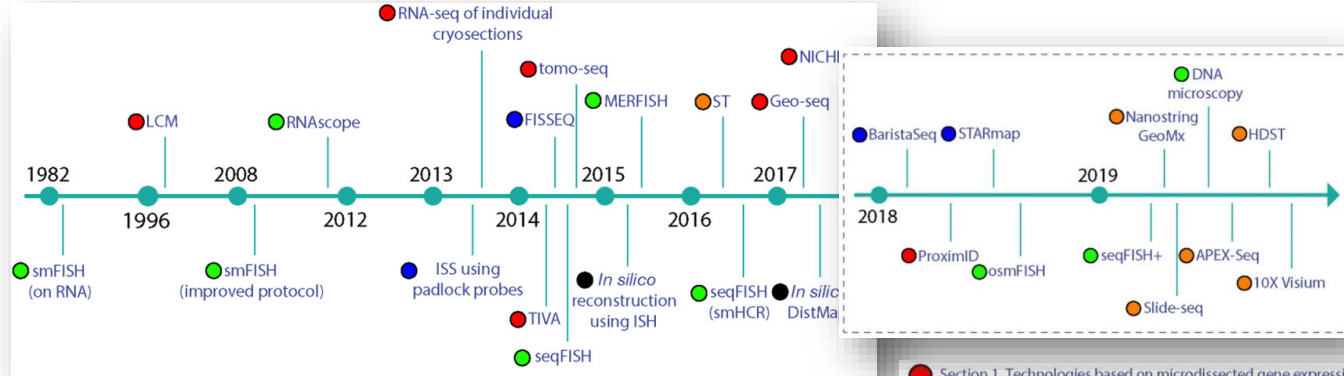
*Nature Method Of the Year 2020*



# Spatial transcriptomics approaches

## Timeline

- Spatial transcriptomics aims at directly visualize gene expression in their original environment,
- It tackles the main limitation of single cell experiment missing the spatial organization,
- A lot of developments in the last years thanks to recent advances in different fields,

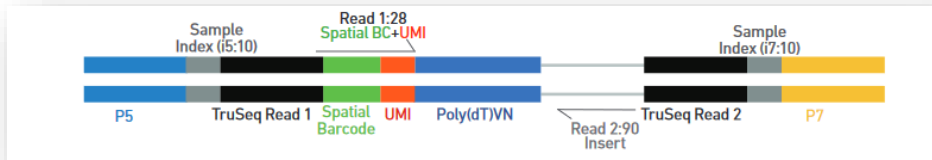
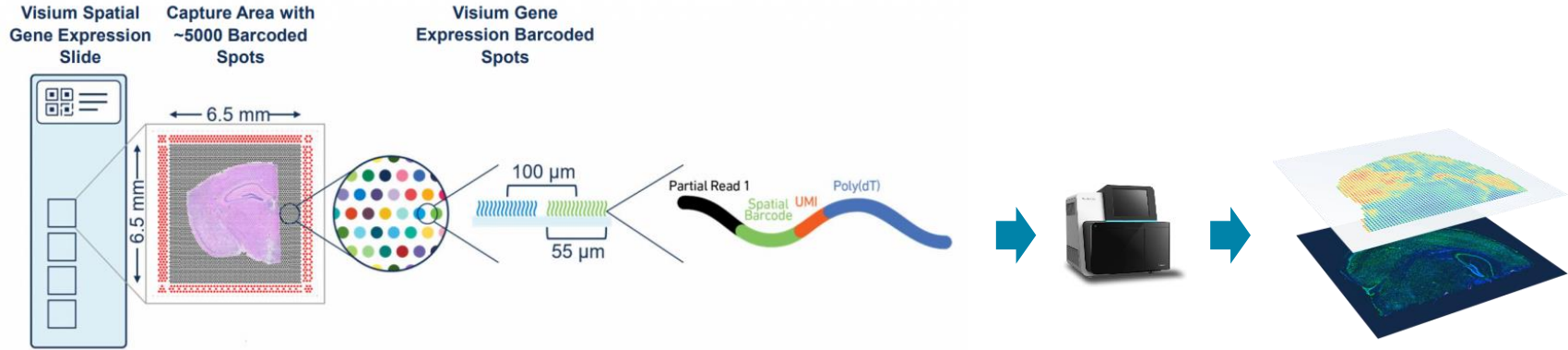


Spatially Resolved Transcriptomes Next Generation Tools for Tissue Exploration  
*Asp et al., BioEssays, 2020*

- Section 1. Technologies based on microdissected gene expression
- Section 2. *In situ* hybridization technologies
- Section 3. *In situ* sequencing technologies
- Section 4. *In situ* capturing technologies
- Section 5. *In silico* reconstruction of spatial data

# In-situ capture spatial transcriptomics

10x Genomics Visium (2019)



→ Spatial barcode / UMI assignment strategy identical to single cell transcriptomics

# Spatial isoform Transcriptomics (SiT)

The spatial landscape of gene expression isoforms in tissue sections

## The spatial landscape of gene expression isoforms in tissue sections

Kevin Lebrigand, Joseph Bergensträhle, Kim Thrane, Annelie Mollbrink, Pascal Barbry, Rainer Waldmann, Joakim Lundeberg

doi: <https://doi.org/10.1101/2020.08.24.252296>



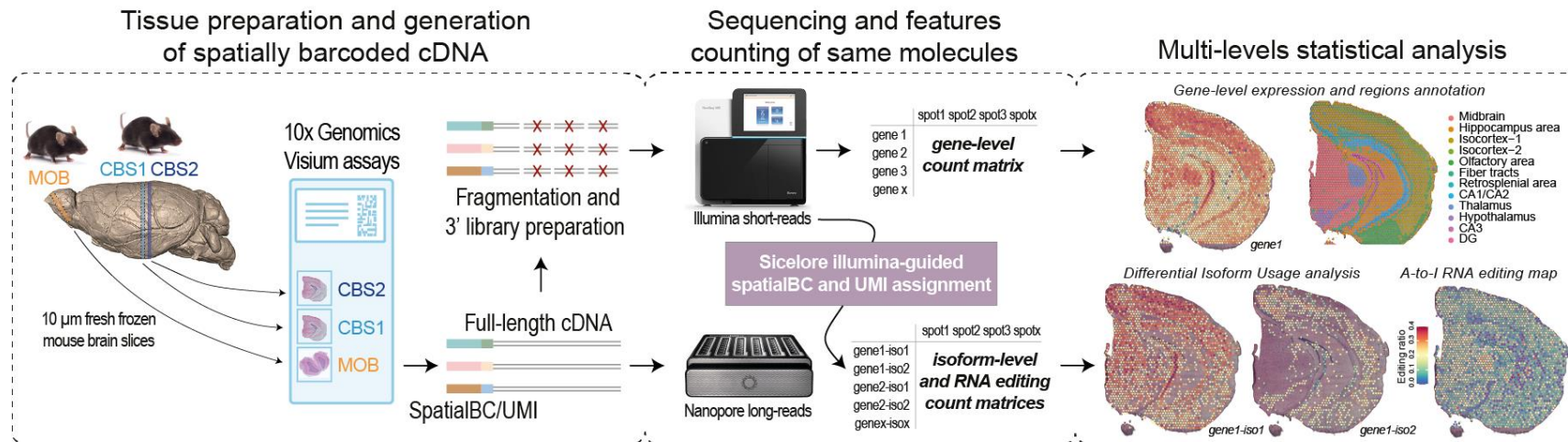
bioRxiv  
THE PREPRINT SERVER FOR BIOLOGY

August 24th, 2020

January 5th, 2022



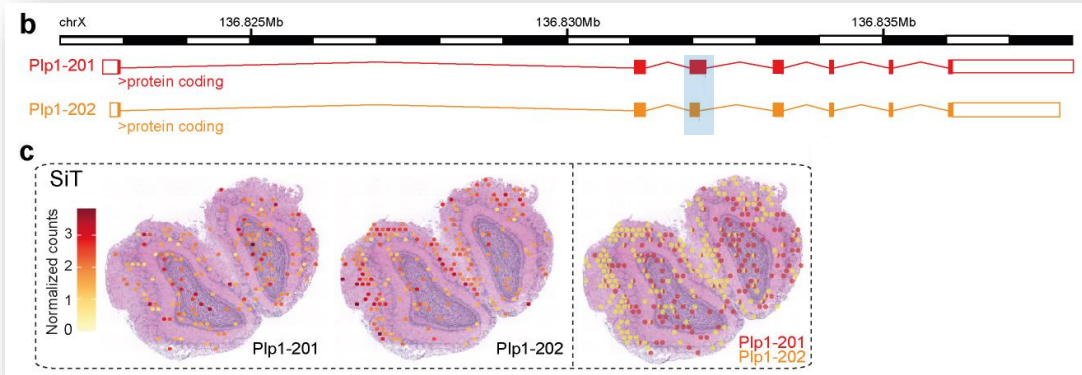
in press





# SiT reveals specific splicing pattern across MOB regions

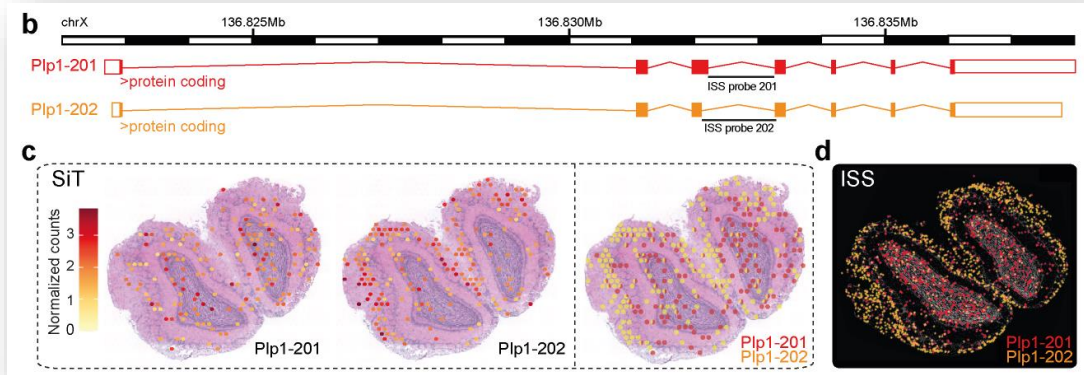
Plp1 Differential Transcript Usage (DTU)



Proteolipid Protein 1 (Plp1) is a gene involved in severe pathologies associated with CNS dysmyelination

# SiT reveals specific splicing pattern across MOB regions

Plp1 Differential Transcript Usage (DTU)



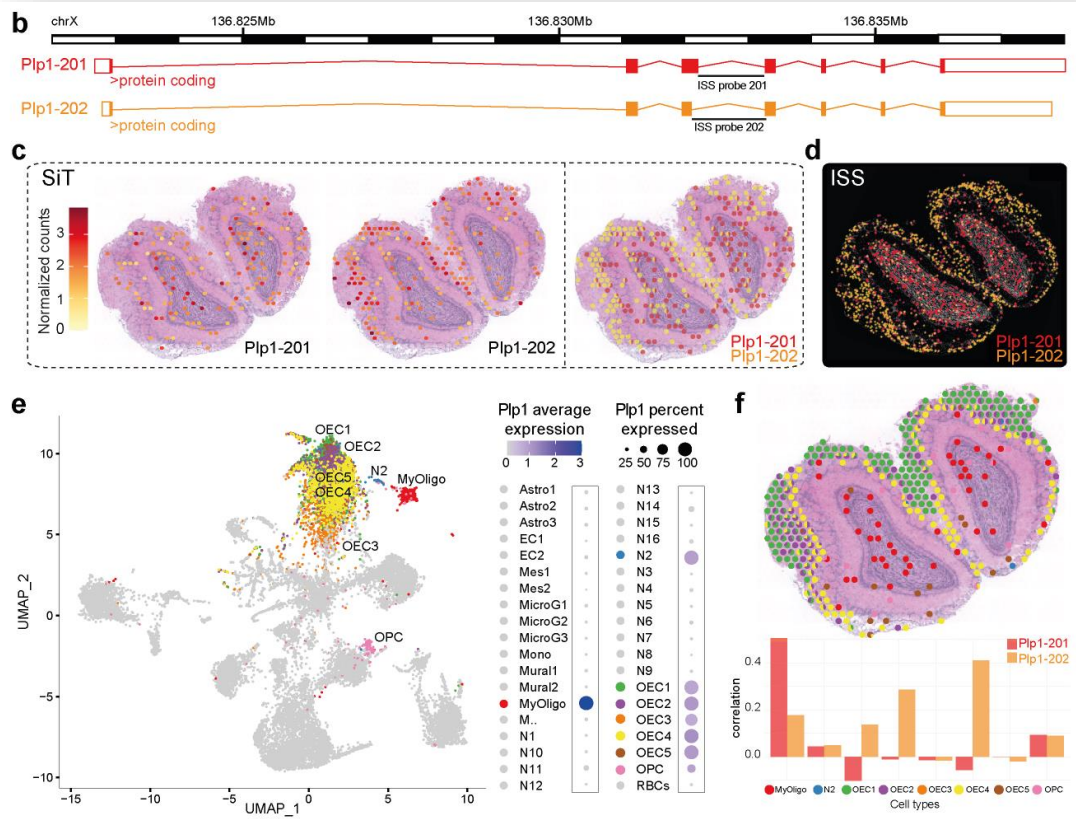
Proteolipid Protein 1 (Plp1) is a gene involved in severe pathologies associated with CNS dysmyelination



In Situ Sequencing Data

# SiT reveals specific splicing pattern across MOB regions

Cell type deconvolution using single cell external dataset (Tepe et al., 2018)



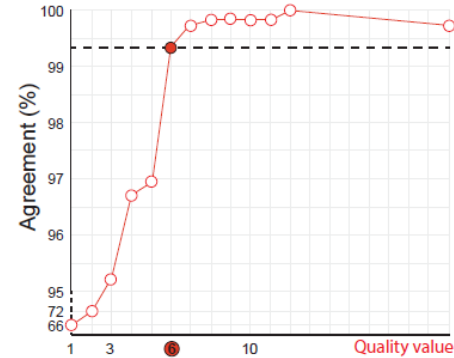
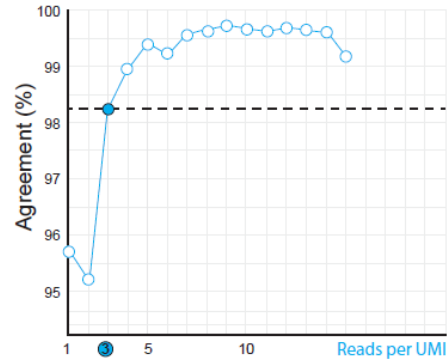
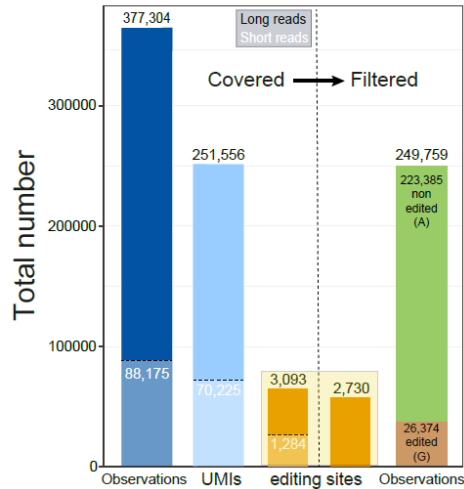
Proteolipid Protein 1 (*Plp1*) is a gene involved in severe pathologies associated with CNS dysmyelination

Spatial spot deconvolution of prominent *Plp1* expresser cell types. Correlation Deconvolution score / *Plp1* isoforms expression correlation shows that *Plp1* is predominantly expressed as *Plp1*-202 by **olfactory ensheathing cells** (OEC) in the **ONL** and as *Plp1*-201 isoform by **myelinating-oligodendrocytes** (MyOligo) in the **GCL**.

# SiT reveals full-length sequence heterogeneity

High confidence SNV call (>99%) calibration using long- vs short-read agreement

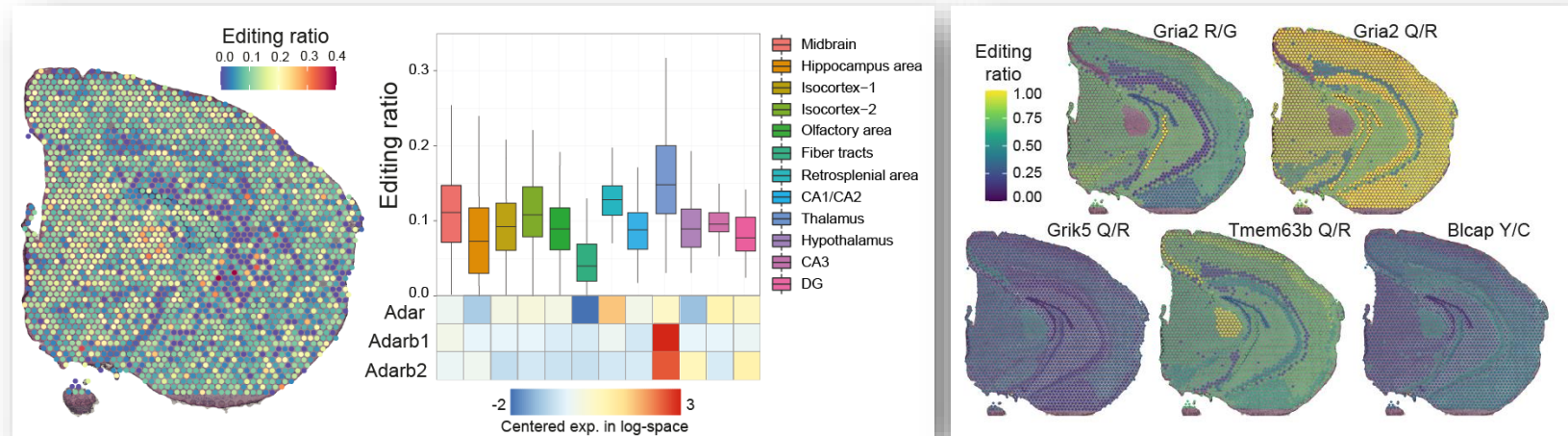
- Exploration of 5,817 A-to-I RNA editing sites described in the literature (Ramaswami et al., 2013 (RADAR), Licht et al., 2019)
- Calibration by looking at agreement between long and short read base calls for 88,175 shared UMI / Editing site observations
  - number of reads per UMI  $\geq 3$
  - consensus Phred score QV  $\geq 6$



# SiT reveals full-length sequence heterogeneity

Mouse brain map of A-to-I RNA editing activity

- Exploration of 5,817 A-to-I RNA editing sites described in the literature (Ramaswami et al., 2013 (RADAR), Licht et al., 2019)



# Single cell and Spatial isoform transcriptomics

## Summary

---

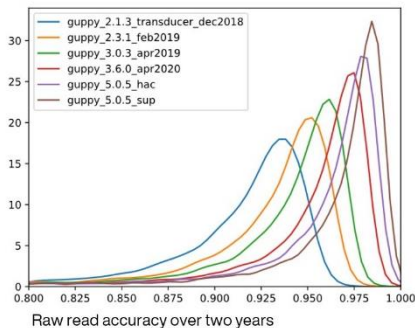
- Accurate single-cell and spatial transcriptomics using Nanopore long-read sequencing is feasible
- Long reads sequencing reveals transcript diversity that is missed with standard short reads workflows
- Single Nucleotide Variation calls (SNV, editing) in single-cell and in a spatial context can be achieved

# Single cell and Spatial isoform transcriptomics

## Summary

---

- Accurate single-cell and spatial transcriptomics using Nanopore long-read sequencing is feasible
- Long reads sequencing reveals transcript diversity that is missed with standard short reads workflows
- Single Nucleotide Variation calls (SNV, editing) in single-cell and in a spatial context can be achieved
- **Sicelore-2.1** : we don't need short reads anymore



Nanopore PromethION sequencing

2018: **20M** reads/FC, **92%** raw read accuracy

2022: **150M** reads/FC, **98%** raw read accuracy



<https://github.com/ucagenomix/sicelore-2.1>

- Visium and single-cell 3' and 5' libraries
- Illumina-free profiling available

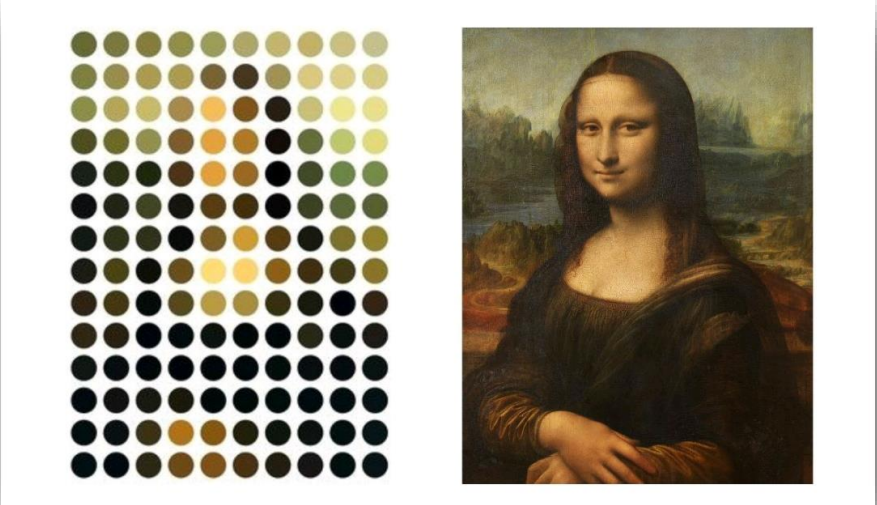
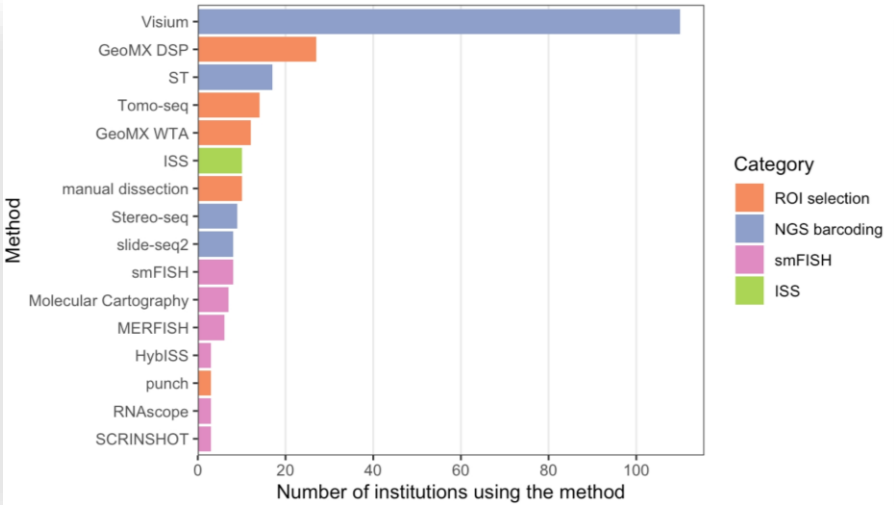
# 03

**Spatial single-cell transcriptomics (imaging)**



# Spatial transcriptomics technologies (2020-2022)

Visium is widely adopted by academics



But is not the ideal readout for spatial biology  
(Akoya credit rough caricature)

# Spatial imaging technologies (2023)

No more sequencing for direct single-cell resolution



**Nanostring CosMx**  
*ISH-based*

- 960 targets
- Sensitivity : < 30-80%
- Resolution: 200 nm
- Imaging area: 16 mm2



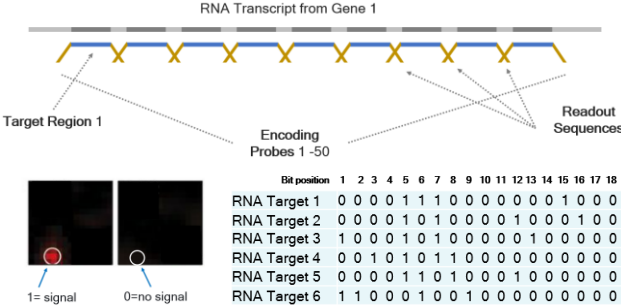
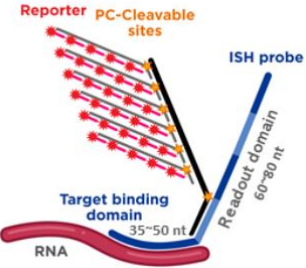
**Vizgen Merscope**  
*Merfish*

- 500 targets (1,000 soon)
- Sensitivity : 30-80%
- Resolution: 100 nm
- Imaging area: 100 mm2



**10xGenomics Xenium**  
*Cartana ISS, padlock probes / RCA*

- 400 targets
- Sensitivity : 5-30%
- Resolution: 200 nm
- Imaging area: 25 mm2



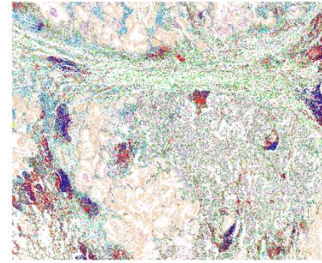
# Spatial imaging technologies (2023)

Compare available datasets

## Vizgen Merscope

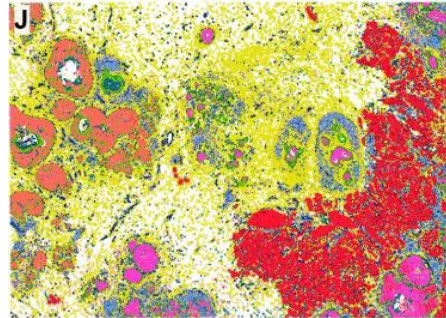
- [Xiaowei Zhuang's lab merfish publications](#)
  - Chen et al., Science (2015)
  - Moffitt et al., PNAS (2016), Science (2018)
  - Emanuel G et al., Nature Methods (2017)
  - Xia C. et al., PNAS (2019, Scientific Reports (2019)
  - Zhang M. et al., Nature (2021)
  - ...
- [Internal data release program](#)
  - Human Immuno-oncology (**breast**, colon, **lung**, liver, skin, prostate, uterine and ovarian) 500 genes, >4 billion transcripts, 9 million cells
  - Mouse Liver Map (347genes)
  - Mouse brain Receptor Map (483 genes)
- [External labs publications](#)
  - Dixon E. et al., Kidney Int. (2022): Kidney
  - Wang et al., Nat. Neuro. (2022): Mouse olfactory Glomerular map
  - Stogsdill et al., Nature (2022): Neocortex microglia
  - ...

## Nanostring CosMx



- Release date: 11/2021
- FFPE Human NSCLC (**Lung**)
- 960 gene targets
- 8 sections for 800k cells
- Imaging area: 8 x 16 mm<sup>2</sup>
- 259,604,214 transcripts
- Mean transcripts/cell: 265

## 10xGenomics Xenium



- Release date: 10/2022
- FFPE Human **Breast** cancer
- 313 gene targets
- 167,885 cells,
- 36,944,521 transcripts
- Imaging area: 40 mm<sup>2</sup>
- Mean transcripts/cell: 193

# Spatial imaging technologies comparison

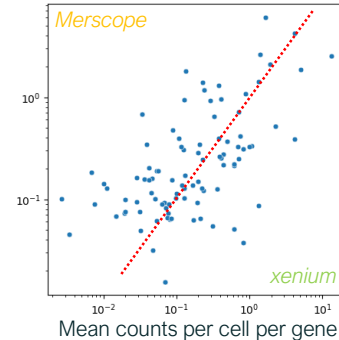
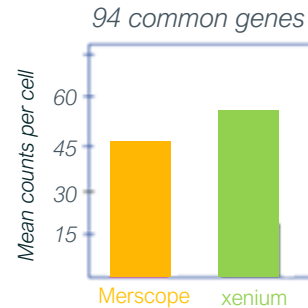
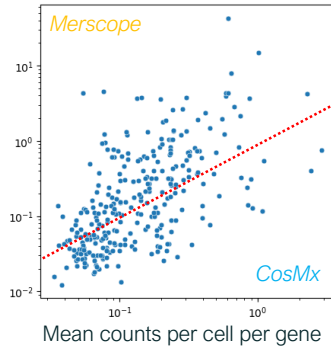
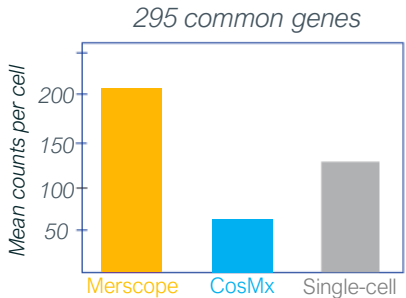
Compare available datasets: Lung and Breast cancer samples



FFPE Human Lung Cancer	Merscope	CosMx
Total cells	353 k (x4)	92 k
Detected transcripts	107 M (x4)	26 M
Gene targets	500	960 (x2)
Total RPKM	9,204	61,680 (x6)
Mean transcripts/cell	302	284



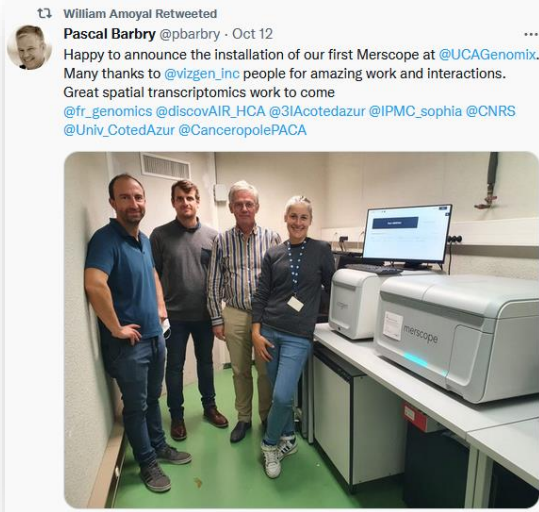
FFPE Human Breast Cancer	Merscope	Xenium
Total cells	713 k (x4)	168 k
Detected transcripts	353 M (x10)	32 M
Gene targets	500	313
Total RPKM	9,909	7,912
Mean transcripts/cell	495	193



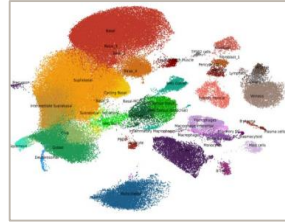
<https://vizgen.com/wp-content/uploads/2022/12/Vizgen-Spatial-Genomics-Data-Quality-eBook-1.pdf>

# MERSCOPE @ UCAGenomiX (Nice-Sophia-Antipolis)

October 2022



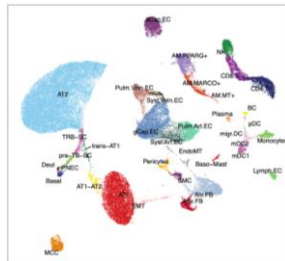
- Human Lung Cell Atlas (CZI)  **discovAIR**  
Discovering the Cellular Landscape of the Airways and Lung Tissue



- 12 control / 2 IPF / 10 COPD patients
- 415k cells (117 samples)
- 48 cell types



- DSRemodeling (mouse model brain, Massimo Mantegazza, IPMC)  
Molecular and functional remodeling in the developmental and epileptic encephalopathy Dravet Syndrome (DS)
- Pulmonary Arterial Hypertension (Christophe Guignabert, Paris-Saclay)



- 7 control / 7 PAH patients
- 69,949 cells
- 39 cell types



# Acknowledgments

## Institut de Pharmacologie Moléculaire et Cellulaire



Pascal Barbry Lab (IPMC, CNRS, France)

- Rainer Waldmann

Joakim Lundeberg Lab (KTH Royal Institute of Technology, Sweden)

- Joseph Bergenstråhle
- Kim Thrane

UCAGenomiX platform (IPMC, CNRS, France)

- Marie-Jeanne Arguel

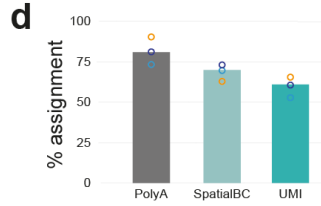
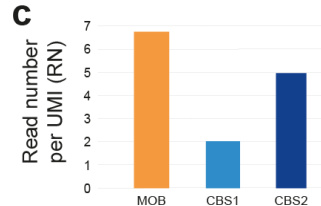
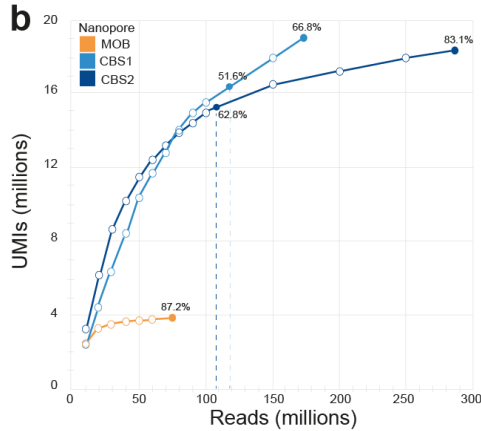




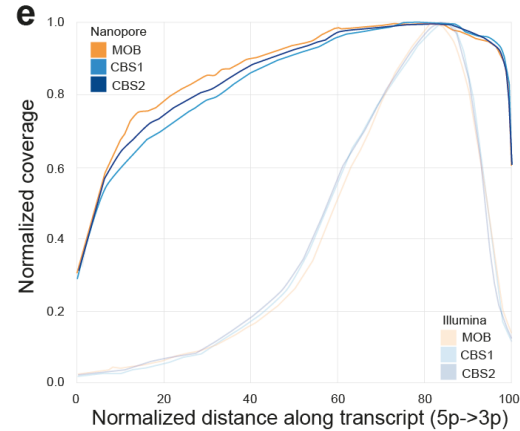
# Spatial isoform Transcriptomics (SiT)

Nanopore long-read sequencing

Sequencing saturation curves per samples



Transcripts full-length coverage



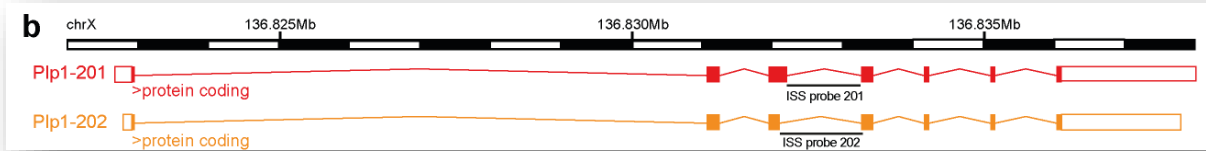
Reads	MOB				CBS1				CBS2								Total	%age	
Date	18 feb. 20	20 mar. 20	18 feb. 20	20 mar. 20	24 feb. 21	12 may 20	13 may 20	19 may 20	25 may 20	25 may 20	26 may 20	27 may 20	09 feb. 21						
Flow cells	PAE06474	PAE59649	PAE01745	PAE59645	PAG52067	PAE59606	PAE59231	PAE32756	PAE32753	PAE31188	PAE21339	PAD99555	PAG56368	13					
Total reads (fastq_pass)	27628000	47272000	24980000	31736000	117280000	22897702	30405384	27492770	18534938	31506774	19108718	25596387	110916000	535354673	79.96	of Total passed reads			
PolyA and Adapter found reads	21318117	47970311	17980183	27286678	80516212	18536047	25199992	22871198	16088962	26777546	15983663	21682530	85837208	428048647	68.90	of PolyA found reads			
SpatialBC found reads	14506264	29316718	12554655	19051597	54323311	14613934	19867830	14666481	11403706	19099469	11266930	14090779	60154119	294915793	59.61	of SpatialBC found reads			
UMIs found reads	10445006	19328468	7323748	10517081	27584331	8616415	11714126	9347072	7557944	12657620	7448718	9031708	34225619	175797856					

CBS1: One flow cell, 117 M reads → 51.6% sequencing saturation  
 CBS2: One flow cell, 111 M reads → 62.2% sequencing saturation  
 → 1 or 2 Promethion flow cells per slice - R9.4, LSK-110 chemistry (2021)



# Spatial imaging technologies

No more sequencing not compatible with isoform transcriptomics



- Validating one exon-exon junction is not really the same as validating a full-length isoform (complete exons layout required)
- Current limitations in # target genes is even more an issue when dealing with the huge amount of exon-exon junctions, Human Ensembl GRCh38.108:
  - 20,036 protein coding genes,
  - 170,185 described isoforms,
  - 626,750 unique exons,
  - 616,988 unique exon-exon junctions
- Even in case of no more limitation in # targets, we would not be able to link together individual exon-exon signal to reconstitute unambiguously each gene isoforms expression

# From Gene-level to Isoform-level long-read profiles

Enhanced description of single-cell transcriptome

Gene-level matrix

	cells																				
Ctla	5	4	6	7	1	4	3	9	5	5	1	4	3	6	4	7	5	2	4		
My16	5	2	5	1	2	4	13	2	5	2	4	4	1	4	8	4	2	3	3	1	1
Pkm	3	2	1	.	.	8	3	9	4	10	5	1	.	4	12	5	1	3	9	1	7
Tecr	3	1	5	2	4	6	5	9	6	3	2	2	1	4	7	1	1	1	3	2	8
Meis2	8	29	3	.	24	6	6	21	25	16	6	1	.	2	29	.	.	6	18	.	10
...																					

Genes

Isoform-level matrix

	cells																					
Ctla..ENSMUST00000107851.9	1	.	.	.	.	.	.	1	.	.	.	.	.	.	.	.	.	.	.	.	.	.
Ctla..ENSMUST00000107845.3	.	1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
Ctla..ENSMUST00000170241.7	3	2	.	.	1	2	.	1	.	.	.	.	.	.	.	.	.	.	.	.	.	.
Ctla..ENSMUST00000107849.9	1	.	3	5	.	3	.	5	3	2	.	1	3	3	4	4	.	3	4	2	3	
Ctla..ENSMUST00000107847.9	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
My16..ENSMUST00000217733.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
My16..ENSMUST00000218127.1	1	.	1	.	.	1	8	.	.	.	.	.	.	.	1	1	.	.	.	.	.	
My16..ENSMUST00000217776.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
My16..ENSMUST00000219236.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
My16..ENSMUST00000164181.1	4	1	2	1	1	2	1	1	3	2	3	2	.	5	3	2	1	2	1	.	.	
My16..ENSMUST00000219554.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	1	.	.	.	.	.	.	.
My16..ENSMUST00000218170.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
My16..ENSMUST00000220307.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
My16..ENSMUST00000217969.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	1	.	.	.	.	.	.	.
Pkm..ENSMUST00000214571.1	.	.	.	.	.	.	.	.	.	.	.	.	.	.	1	.	.	.	.	.	.	.
Pkm..ENSMUST00000163694.3	2	2	.	.	3	.	7	2	6	2	.	2	9	1	1	1	3	1	.	.	.	
Pkm..ENSMUST0000034834.15	1	.	1	.	3	3	.	1	.	.	.	1	.	.	1	.	1	2	.	3	.	.
Meis2..ENSMUST0000028639.12	.	.	.	.	.	.	.	.	.	.	.	.	.	.	1	.	.	.	.	.	.	.
Meis2..ENSMUST00000189640.6	.	.	.	.	.	.	.	.	.	.	.	.	.	.	1	.	.	.	.	.	.	.
Meis2..ENSMUST00000110908.8	.	.	.	.	2	.	.	1	.	1	.	.	.	.	1	.	.	.	.	.	.	.
Meis2..ENSMUST00000110907.7	.	.	.	.	1	.	.	1	.	1	.	.	.	.	1	.	.	.	.	.	.	.
Meis2..ENSMUST00000110906.8	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
Meis2..ENSMUST00000074285.7	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.	.
Meis2..ENSMUST00000102538.10	.	.	.	.	.	.	.	.	.	.	.	.	.	.	1	1	.	.	.	.	.	.
...																						

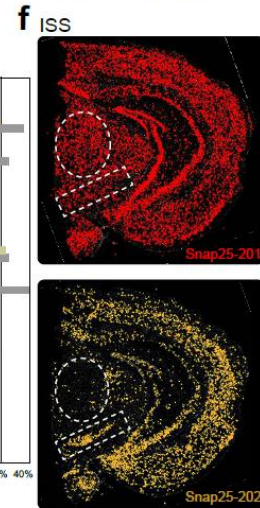
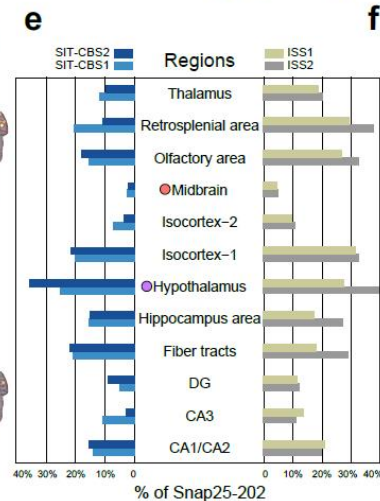
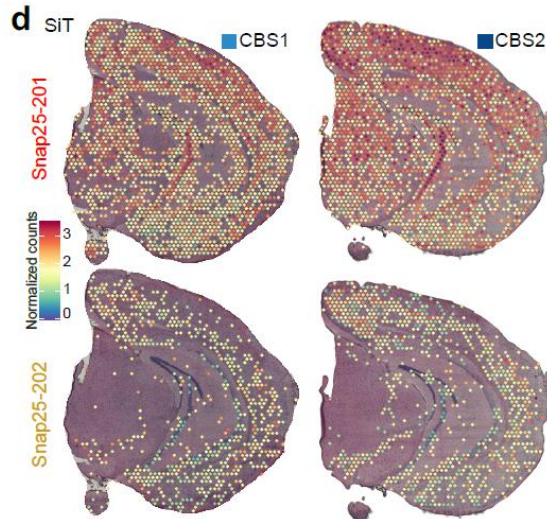
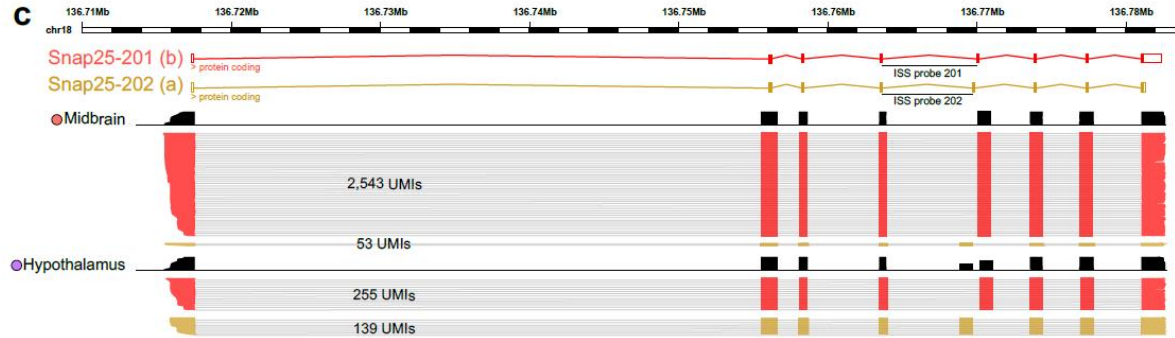
Isoforms

Isoform-allelic-level matrix

Actually in development

# Snap25 DTU across CBS regions

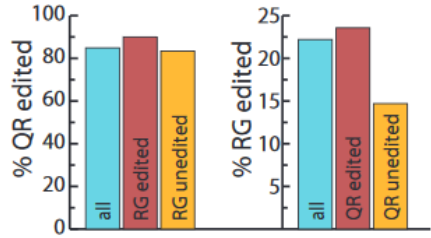
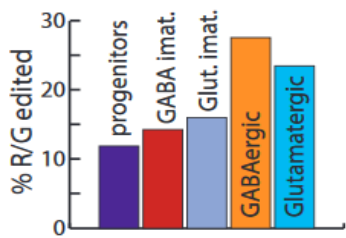
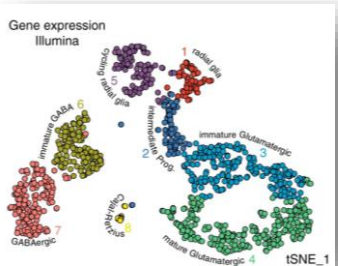
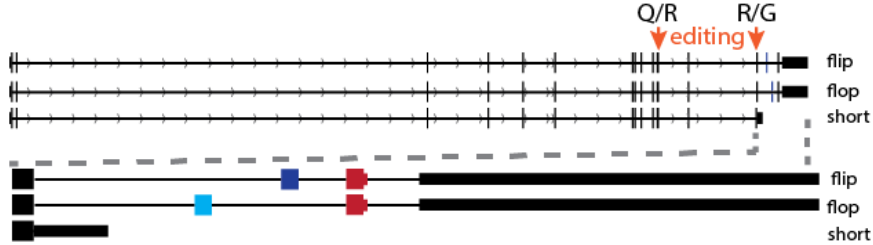
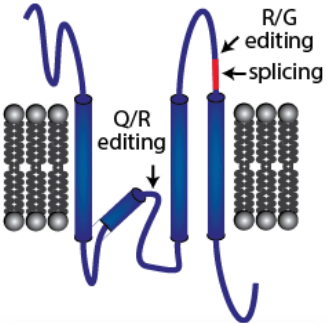
Presynaptic plasma membrane protein involved in the regulation of neurotransmitter release



# Single-cell long-read transcriptomics reveals sequence heterogeneity

RNA editing of the AMPA receptor Gria2

Q/R site regulates AMPA receptor  $Ca^{2+}$ -permeability  
 R/G site is involved in desensitization and recovery of the receptor

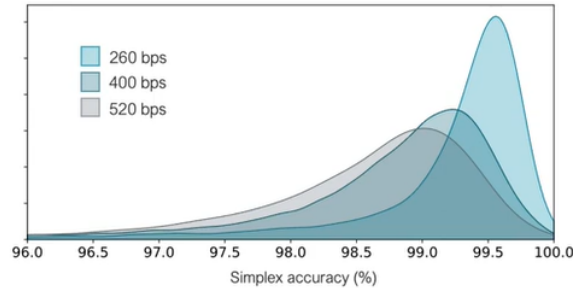


## Nanopore Chemistry

### Tuneable run settings

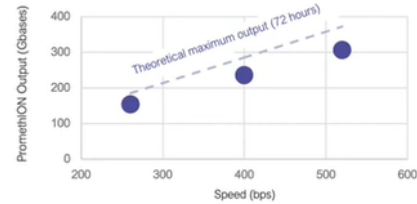
#### Same chemistry, different conditions

- Depending on the application requirement R10.4 / Kit14 can be run at different temperatures to tune the accuracy vs output
- Appropriate models will be chosen automatically or selected by user
- Example speeds and outputs
  - 260, 400, 520 bps
- Different accuracies
  - Simplex: 99.6%, 99.2%, 99.0%
  - Duplex: Q31, Q30, Q29



#### Choice of run condition

- Current focus is the 400 bps default condition
  - Demonstrated 236 Gbases from PromethION flow cell
  - Modal accuracy 99.2% (Q21)
- Choice of run condition coming soon
  - "Accuracy" 260 bps, "Default" 400 bps, "Output" 520 bps
  - 99.6% (Q24) @ 260 bps, 307 Gbases @ 520 bps



R9 series



R10 series



Rx series



# London Calling 2022



## In your hands from \$1,000

Scalability and accessibility



Flongle

MinION

MinION Mk 1c

GridION

PromethION P2 solo  
(in EA)

PromethION P2  
(in dev)

PromethION 24  
With A100 compute

PromethION 48  
With A100 compute

<b>\$1,640</b>	<b>\$1,000</b>	<b>\$4,900</b>	<b>\$49,955</b>	<b>\$10,455</b>	<b>\$59,995</b>	<b>\$225,000</b>	<b>\$310,000</b>
• 12 flow cells	• 1 flow cell	• 6 flow cells	• 60 flow cells	• 8 flow cells	• 48 flow cells	• 192 flow cells	• 288 flow cells
• Flongle adapter	• 1 kit	• 1 kit	• 10 kits	• 2 kits	• 8 kits	• 42 kits	• 48 kits
Use with GridION or MinION		• 12M SW licence	• 12M SW licence	• 12M SW licence	• 12M SW licence	• 12M SW licence	• 12M SW licence

Same sample prep – same sequencing chemistry – same data analysis

## Technical Concept

